

**In the specification:**

Please replace the paragraph at page 40, lines 12-21 with the following:

Genomic DNA was extracted from tomato leaves (20g) according to the procedure of Bedbrook (1981). 15 µg of purified DNA were cut with EcoRI, BamHI, HindIII, XbaI, and HaeIII (all enzymes from Boehringer Mannheim), respectively, and electrophorized on a 0.8% TAE agarose gel (Sambrook et al. 1989). After the denaturation and renaturation of the agarose gel, the DNA was transferred (Vacuum Blotter, Appligene) to positively charged Nylon Plus membrane (Qiagen), and finally UV<sub>312nm</sub> crosslinked (0.3J/cm<sup>2</sup>). Hybridization of the Southern blot against random primed <sup>32</sup>P-labeled RdRP24 DNA (see above) was performed as described by Amasino (1986). Briefly, blots were hybridized at 42°C in 0.25 M NaHPO<sub>4</sub>, pH 7.2, 0.25 M NaCl, 7% SDS, 1 mM EDTA, 50% formamide with or without 5-20% (w/v) PEG (M<sub>r</sub> 6-7.5 X 10<sup>3</sup>) or DexSO<sub>4</sub> (M<sub>r</sub> 5 X 10<sup>5</sup>) on a rocker platform. Prehybridization was for 5-10 minutes without change of solution. After hybridization, blots were washed once in 2X SSC (1X SSC = 0.15M NaCl, 0.015 M Na Citrate, pH 7.0) for five minutes at room temperature, twice in 0.25 M NaHPO<sub>4</sub>, 2% SDS, 1 mM EDTA for 20-60 minutes at 65°C.